

Genetic Polymorphism of Interleukin-18 Gene Promoter Region in Rheumatoid Arthritis Patients from Southern India

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ABSTRACT

Background: Interleukin-18 (IL-18) is a pro inflammatory cytokine which plays a key role in the acute and chronic inflammatory phases of Rheumatoid Arthritis (RA). The Single Nucleotide Polymorphisms (SNPs) of IL-18 gene promoter region at positions -137 and -607, are postulated to be associated with RA. To test this, this study aimed to identify the association between these SNPs of the IL-18 gene promoter region of RA in south Indian patients.

Materials and Methods: This study was carried on 190 subjects among which 90 were RA patients and 100 were age and sex matched controls. Genomic DNA was extracted by Salting out method. IL 18 gene promoter region SNPs, IL 18 - 607 and IL 18 -137 were amplified by using sequence specific primers. The amplified products of different samples were separated by using a 1.5% agarose gel, stained with ethidium bromide and

photographed. All statistical analyses were carried out by using SYSTAT 12 software.

Results: At position 607, the frequencies of C allele, CC genotype, A allele and AA genotype were found to be significantly higher in patients and controls respectively and there was no significant difference in CA genotype. At position 137, there was no significant difference between the two groups with regard to G and C alleles but there was a significant increase in GG genotype of patients and CC genotype of controls. There was no association between duration of morning stiffness, rheumatoid factor positivity or negativity, age of onset and gender with distribution of genotypes and alleles.

Conclusion: C allele, CC genotype at position-607 and GG genotype at position-137 are risk factors and A allele, AA genotype at position-607 and CC genotype at position-137 have protective effect for RA.

Keywords: Interleukin-18, Genotype, Pro inflammatory cytokine

INTRODUCTION

Rheumatoid Arthritis (RA) is a chronic, generally progressive, autoimmune disease that causes functional disability, significant pain and joint destruction [1]. In adult Indian population, a prevalence rate of 0.75% was observed [2]. This is a common disease seen in tropical areas [3]. Women are mostly affected by RA. It affects thrice as many women as men [4].

The pathogenesis of RA involves the following stages i.e. initiation, perpetuation and tissue damage. Each stage involves different cell and molecular interactions [5-6]. Interleukin-18 (IL-18) a pro inflammatory cytokine and it plays significant role in the pathogenesis of RA. The gene, IL-18 is mapped to short arm of chromosome 11 (11q22.2-22.3) [7]. IL-18 is found in the synovial tissues and enhanced levels of IL-18 are measured in the joints and sera of RA patients. IL-18 enhances the infiltration of inflammatory cells into the synovial tissues [8]. The role of IL-18 in RA is as follows; in chondrocytes, IL-18 increases gene expression for inducible nitric oxide synthetase, cyclooxygenase 2 and stromelysin [9]. Exposure of human articular cartilage to IL-18 increases the release of glycosaminoglycans, which are by-products of its degradation.

IL-18 protein expression is regulated by the IL-18 promoter gene [10,11] through two single nucleotide polymorphisms (SNPs), at positions -607 and -137 in the promoter region. These promoter regions are expected to be the binding sites for Cyclic (Adenosine 30, 50-cyclic monophosphate) AMP-responsive element-binding protein (CREB) [12] and Human histone H4 gene-specific transcription factor-1(H4TF-1) [13]. At position -607, the alteration from C-(cytosine) to A- (adenosine) nucleotide disrupts a potential

CREB binding site and at position -137, the change from G- (guanine) to C- (cytosine) nucleotide affects the H4TF- 1-binding site. In the IL-18 gene promoter transcription activity assay, following stimulation, low promoter activities were observed for A and C alleles at positions -607 and -137, respectively. In contrast, higher promoter activities were observed for C and G alleles at similar positions [14]. RA patients may have higher IL-18 gene promoter transcription activities as causal factors of their disease pathogenesis. It has been postulated that RA patients would have higher frequencies of C alleles at position -607 and higher frequencies of G alleles at position -137 of the IL-18 promoter gene. Alternatively, higher frequencies of A allele at position -607 and higher frequencies of C allele at position -137 would confer protective effects against the development of RA. To test these postulations, this study aimed to identify the associations between these SNPs of the IL-18 promoter gene region in RA disease, in South Indian patients. In addition to its pro inflammatory activity in RA, IL-18 also contributes to other diseases such as Cancer [15], Crohn's disease [16], Type 1 Diabetes [17,18] and Adult-Onset Still's disease [19].

MATERIALS AND METHODS

This study was carried out on 190 subjects, of which 90 (65 females and 25 males) were RA patients and 100 were age and sex matched controls. The laboratory work was done in Department of Human Genetics, Andhra University and Pediatric Research and Genetic Lab, Maulana Azad Medical College, New Delhi, India. All patients fulfilled the American College of Rheumatology (ACR) 1987 revised criteria for classification of RA [20] and had disease history of minimum three years. Five ml intravenous blood samples

were collected from patients and controls into EDTA vacutainers under aseptic conditions after obtaining informed consents from subjects.

PCR Amplification

Genomic DNA was extracted by Salting out method [21] and it was quantified by using a spectrophotometer. An absorbance ratio of 1.8:2.0 or greater was considered and the final solution was stored at -4°C. The set of sequence specific primers, as was illustrated in previous studies [22] was used to amplify the target DNA in the promoter region of IL-18 -137 and -607 and it has been summarized in [Table/Fig-1].

Amplification of IL-18 -607

The Hot start period was performed for 2 minutes at 94°C, followed by 94°C - 20 seconds, 64°C - 40 seconds, 72°C - 40 seconds for 7 cycles and 94°C - 20, seconds 57°C - 40 seconds, 24 cycles, 72°C - 40 seconds and 72°C - 5 minutes.

IL18-607-F1	5'-GTTGCAGAAAGTGAAAAATTATTAC-3'
IL18-607-F2	5'-GTTGCAGAAAGTGAAAAATTATTAA-3'
IL18-607-R	5'-TAACCTCATTGAGACTTCC-3'
IL18-607-CF	5'-CTTTGCTATCATTCCAGGAA-3'
IL18-137-F1	5'-CCCCAACTTTTACGGAAGAAAAG-3'
IL18-137-F2	5'-CCCCAACTTTTACGGAAGAAAAC-3'
IL18-137-R	5'-AGGAGGGCAAATG CACTGG-3'
IL18-137-CF	5'-CCAATAGGACTGATTAT TCCGCA-3'

[Table/Fig-1]: IL-18 promoter region gene primers (Sivalingam et al., 2003)

IL-18 137 Amplification

The Hot start period is for 2 minutes at 94°C, followed by 94°C - 20 seconds, 68°C - 60 seconds for 5 cycles and 72°C - 40 seconds, 94°C - 20 seconds, 62°C - 40 seconds, 25 cycles and 72°C - 40 seconds, 72°C - 5 minutes. An amplification product of 196 bp was detected for position -607 and a 261 bp product was detected for position -137. The amplified products for different samples were separated by using a 1.5% agarose gel, stained with ethidium bromide and photographed.

STATISTICAL ANALYSIS

All statistical analyses were done by using SYSTAT 12 software. A two-sided t-test was used to compare continuous variables like Haemoglobin %, ESR and cell counts (total and differential) between rheumatoid factor (+ve) and rheumatoid factor (-ve) patients. The genotypic, allelic frequencies of subjects and the genotypic and allele frequencies with respect to gender, age of onset and clinical features (morning stiffness and rheumatoid factor) were compared by using z-test for comparison of proportions. In all the above statistical tests, a probability value of <0.05 was considered as statistically significant.

RESULTS

The demographic and clinical factors have been summarized in [Table/Fig-2]. The female to male ratio was 2.6:1, mean disease duration was 5±5 years and the age of onset was 39±12 years. When the clinical profiles were compared between rheumatoid factor (+ve) and rheumatoid factor (-ve) patients, they showed no association between them [Table/Fig-3].

The alleles and genotype frequencies of IL 18 -607 have been presented in [Table/Fig-4,5]. At position 607, the frequencies of C allele, CC genotype, A allele and AA genotype were found to be significantly higher in patients and controls respectively. The difference was statistically significant (p=0.0000) and no significant difference was seen in CA genotype.

The allelic and genotypic frequencies of IL 18 -137 have been presented in [Table/Fig-6,7]. At position 137, no significant difference was seen between the two groups with regard to G and C alleles, but significant increases were observed in GG genotype of patients and CC genotype of controls (p=0.0000).

The distributions of genotypic and allelic frequencies of IL 18-607 and IL 18-137 in relation to duration of morning stiffness, rheumatoid factor (+ve) and rheumatoid factor (-ve), age of onset and gender have been given in [Table/Fig-8,9]. No significant association was seen between these parameters and distributions of genotypes and alleles of IL 18-607 and IL 18-137 polymorphism in RA [Table/Fig-10,11].

No	Parameters	Control	RA
1	Number of patients	100	90
1.1	Females	50	65
1.2	Males	50	25
1.3	Female to Male ratio (F:M)	1:1	2.6:1
2	Age (years)	48±12	46 ± 12
3	Age of onset (years)	NA	39 ± 12
4	Disease duration (years)	NA	5 ± 5
5	Rheumatoid Factor positive cases	NA	47/87 (54.02%)
5.1	Females	NA	34/64 (53.12%)
5.2	Males	NA	13/23 (56.52%)
6	Erythrocyte sedimentation rate (ESR)	NA	41 (6-110)
7	C-reactive protein (CRP) positive cases	NA	24/87 (23%)
7.1	Females	NA	16/64 (25%)
7.2	Males	NA	5/23 (21.73%)

[Table/Fig-2]: Demographic and Clinical factors of rheumatoid arthritis patients

Parameter	RF+ve (Mean±SD)	RF-ve (Mean±SD)	p-value	Significance	Sd
HB (%)	11 ± 2	11 ± 2	0.12	NS	2.96
ESR (cm/hr)	39 ± 18	44 ± 23	0.20	NS	2.2
Total count		9004 ± 1921	0.75	NS	2.96
	Neutrophils	65 ± 8	66 ± 7	0.78	NS
Differential count	Lymphocytes	29 ± 8	29 ± 7	0.93	NS
	Eosinophils	4 ± 2	3 ± 2	0.26	NS
	Monocytes	2 ± 1	2 ± 1	0.30	NS

[Table/Fig-3]: Comparison of clinical profiles in sero positive and sero negative rheumatoid arthritis patients

RF+ve= rheumatoid factor positive, RF-ve= rheumatoid factor negative, p= p-value – probability value of the statistical test, SD= standard deviation, S= significant, NS= not significant

Alleles	RA Patients		Controls		p-value	Significance
	n (160)	Frequency	n (200)	Frequency		
A	41	0.26	128	0.64	0.000	S
C	119	0.74	72	0.36	0.000	S

[Table/Fig-4]: Allelic frequencies of IL-18 promoter gene at position -607 in rheumatoid arthritis patients and controls

n = number of individuals, p-value – probability value of the statistical test, S = significant, NS= not significant

Genotypes	RA Patients		Controls		p-value	Significance
	n (80)	Frequency	n (100)	Frequency		
CC	42	0.52	10	0.10	0.000	S
CA	35	0.44	52	0.52	0.271	NS
AA	3	0.04	38	0.38	0.000	S

[Table/Fig-5]: Genotypic frequencies of IL-18 promoter gene at position -607 in rheumatoid arthritis patients and controls

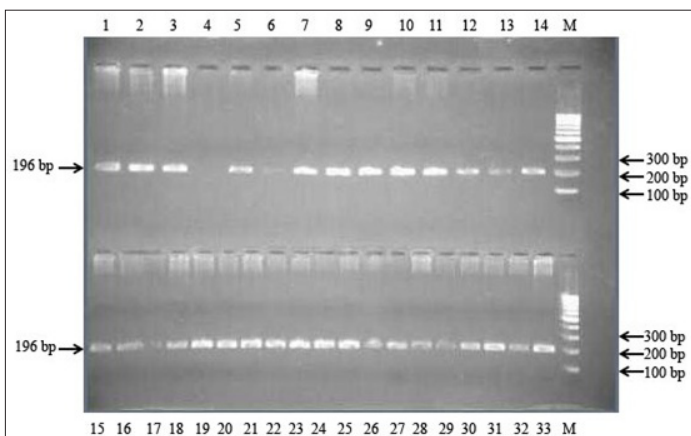
n – number of individuals; p-value – probability value of the statistical test, S= significant, NS= not significant

Alleles	RA Patients		Controls		p-value	Significance
	n (140)	Frequency	n (200)	Frequency		
G	103	0.74	141	0.70	0.536	NS
C	37	0.26	59	0.30	0.536	NS

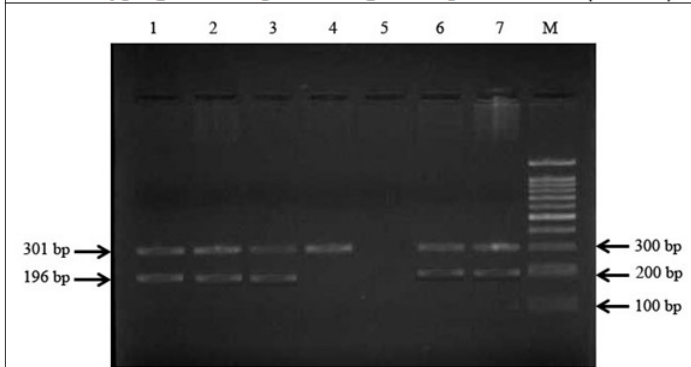
[Table/Fig-6]: Allelic frequencies of IL-18 promoter gene at position -137 in rheumatoid arthritis patients and controls
 n = number of alleles, p-value = probability value of the statistical test, S= significant, NS= not significant

Genotypes	RA Patients		Controls		p-value	Significance
	n (70)	Frequency	n (100)	Frequency		
GG	42	0.60	20	0.20	0.000	S
GC	19	0.27	19	0.19	0.209	NS
CC	9	0.13	61	0.61	0.000	S

[Table/Fig-7]: Genotypic frequencies of IL-18 promoter gene at position -137 in rheumatoid arthritis patients and controls
 n – number of individuals; p-value – probability value of the statistical test, S= significant, NS= not significant

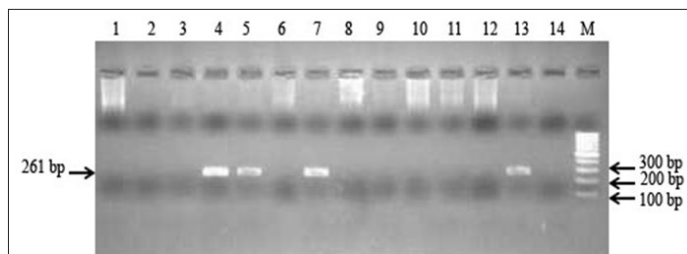


Genotyping of IL-18 promoter gene at position 607 (C allele)

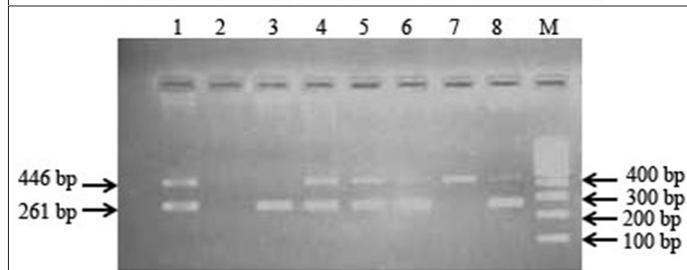


Genotyping of IL-18 promoter gene at position 607 (A allele)

[Table/Fig-8]: Genotyping of IL-18 promoter gene at position -607. A 1.5% agarose gel stained with ethidium bromide after PCR amplification for the position -607 genotyping. Amplification products of 196-bp for the alleles C and A (arrow) were detected. The 301-bp band corresponds to the PCR internal control. The right lane contains a 100-bp marker (M)



Genotyping of IL-18 promoter gene at position 137 (C allele)



Genotyping of IL-18 promoter gene at position 137 (G allele)

[Table/Fig-9]: Genotyping of IL-18 promoter gene at position -137: A 2% agarose gel stained with ethidium bromide after PCR amplification for the position -137 genotyping. Amplification products of 261-bp for the alleles G and C were detected. The 446-bp band corresponds to the PCR internal control. The right lane contains a 100-bp marker (M)

DISCUSSION

The studies done on IL-18 in RA are conflicting and inconsistent. An earlier report made by Sivalingam et al., [22] on RA patients studied in Singapore showed similar results for position -607 i.e. high frequencies of A and C alleles among controls and patients respectively, but they were statistically insignificant. Regarding genotypic frequencies, genotype AA was found to be significantly higher in controls and no statistical significance was found for CA and CC genotypes. The allelic or genotypic frequencies seen at position -137 between subjects were also insignificant. Rueda et al., [23] and Pawlik et al., [24] reported that, IL-18-137 and -607 promoter polymorphisms were not significant with respect to RA courses and severities. Pan et al., [25] reported that IL-18 gene promoter -607 A/C polymorphism was not associated with development of autoimmune diseases. Ying et al., [26] reported that the genotype and allele frequency of IL-18-607 were not associated with IL-18 serum levels. Huang et al., [27] reported that IL-18-607 polymorphism was associated with RA, but not with IL-18-137 polymorphisms. Gracie et al., [28] reported that both SNPs found at positions -137 and -607 were involved in pathogenesis of RA. When other auto immune diseases were considered, the results were found to be very conflicting. Takada et al., [29] reported that C allele, at position -607, was a risk factor for sarcoidosis in the Japanese population. Zhou et al., [30] reported that it was unlikely for these SNPs to confer susceptibility to sarcoidosis [Table/Fig-10,11].

Geno type / allele	Morning Stiffness						Rheumatoid Factor						Age of onset						Gender					
	<1 (n=46)		>1 (n=18)		p-value	Significance	Positive (n=34)		Negative (n=30)		p-value	Significance	>35 years (n=47)		<35 years (n=25)		p-value	Significance	Females (n=46)		Males (n=19)		p-value	Significance
	n	%	n	%			n	%	n	%			n	%	n	%			n	%	n	%		
	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%		
AA	1	2.17	1	5.55	0.484	NS	1	2.94	1	3.33	0.928	NS	6	12.76	4	16.0	0.705	NS	2	4.34	1	5.26	0.872	NS
CA	22	47.82	8	44.44	0.807	NS	13	38.23	17	56.66	0.140	NS	23	48.93	7	28.0	0.086	NS	22	47.28	8	42.10	0.673	NS
CC	23	50.0	9	50.0	1.000	NS	20	58.82	12	40.0	0.132	NS	18	38.29	14	56.0	0.150	NS	22	47.28	10	52.63	0.724	NS
A-allele	24	26.08	10	27.77	0.845	NS	15	22.05	19	31.66	0.219	NS	35	37.23	15	30.0	0.385	NS	26	28.62	10	26.13	0.821	NS
C-allele	68	73.91	26	72.22	0.845	NS	53	77.94	41	68.33	0.219	NS	59	62.76	35	70.0	0.385	NS	66	71.73	28	73.68	0.821	NS

[Table/Fig-10]: Genotype and allele frequencies of IL 18-607 polymorphism in rheumatoid arthritis patients in relation to clinical features
 n – number of patients, <1 =less than one hour, >1 = more than one hour, <35 = less than 35 years, >35 =more than 35 years, p-value = probability value of the statistical test, S = significant, NS = not significant

Geno type / allele	Morning Stiffness				Rheumatoid Factor				Age of onset				Gender											
	<1 (n=45)		>1 (n=18)		Positive (n=34)		Negative (n=29)		>35 years (n=42)		<35 years (n=24)		Females (n=45)		Males (n=23)									
	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%								
GG	27	60.0	11	61.11	0.935	NS	20	58.82	18	62.06	0.792	NS	22	52.38	16	67.66	0.258	NS	27	60.0	11	47.82	0.338	NS
GC	12	26.66	5	27.77	0.928	NS	10	29.41	7	24.13	0.638	NS	12	28.57	5	20.83	0.489	NS	10	22.22	7	30.43	0.459	NS
CC	6	13.33	2	11.11	0.810	NS	4	11.46	4	13.79	0.809	NS	8	19.04	3	12.5	0.492	NS	8	17.77	5	21.73	0.694	NS
G-allele	66	73.33	27	75.0	0.847	NS	50	73.52	43	74.13	0.938	NS	56	66.66	37	77.08	0.207	NS	64	71.11	29	63.04	0.338	NS
C-allele	24	26.66	9	25.0	0.847	NS	18	26.47	15	25.86	0.938	NS	28	33.33	11	22.91	0.207	NS	26	28.88	17	36.95	0.338	NS

[Table/Fig-11]: Genotype and allele frequencies of il 18-137 polymorphism in rheumatoid arthritis patients in relation to clinical features
n = number of patients, <1 =less than one hour, >1 = more than one hour, <35 = less than 35 years, >35 =more than 35 years, p value = probability value of the statistical test, S = significant, NS = not significant

CONCLUSION

On comparing results of the above studies and our findings, it was concluded that C allele and CC genotype at position -607 and GG genotype at position-137 were risk factors for RA. A allele and AA genotype at position-607 and CC genotype at position -137 had protective effects.

Despite these encouraging findings, our study had few limitations. It did not include patients with intermediate and mild severities and also, the number of subjects which was studied was small. By resolving these limitations, this study can be made useful, because the polymorphisms which have been studied here have been linked with progression of the disease. These polymorphisms, therefore, provide a simple, rapid and cost effective tool for prediction of RA. Further, because these polymorphisms do not change over time, they can be used at an early stage of the disease course, in order to individualize therapy and to minimize articular damage.

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